



This form should be used for all taxonomic proposals. Please complete all those modules that are applicable (and then delete the unwanted sections). For guidance, see the notes written in blue and the separate document "Help with completing a taxonomic proposal"

Please try to keep related proposals within a single document; you can copy the modules to create more than one genus within a new family, for example.

MODULE 1: **TITLE, AUTHORS, etc**

Code assigned:	2016.002aS	(to be completed by ICTV officers)
Short title: Create 2 new species (<i>Avisivirus B</i> , <i>Avisivirus C</i>) in the genus <i>Avisivirus</i> (e.g. 6 new species in the genus <i>Zetavirus</i>)		
Modules attached (modules 1 and 11 are required)	2 <input checked="" type="checkbox"/> 3 <input type="checkbox"/> 4 <input type="checkbox"/> 5 <input type="checkbox"/> 6 <input type="checkbox"/> 7 <input type="checkbox"/> 8 <input type="checkbox"/> 9 <input type="checkbox"/> 10 <input type="checkbox"/> 11 <input checked="" type="checkbox"/>	

Author(s):

Roland Zell, Eric Delwart, Alexander E. Gorbalenya, Tapani Hovi, Andrew M.Q. King, Nick J. Knowles, A. Michael Lindberg, Mark A. Pallansch, Ann C. Palmenberg, Gabor Reuter, Peter Simmonds, Tim Skern, Glyn Stanway and Teruo Yamashita

Corresponding author with e-mail address:

Roland Zell (roland.zell@med.uni-jena.de)

List the ICTV study group(s) that have seen this proposal:

A list of study groups and contacts is provided at <http://www.ictvonline.org/subcommittees.asp> . If in doubt, contact the appropriate subcommittee chair (fungal, invertebrate, plant, prokaryote or vertebrate viruses)

Picornaviridae Study Group

ICTV Study Group comments (if any) and response of the proposer:

Date first submitted to ICTV:

15/06/2016

Date of this revision (if different to above):

ICTV-EC comments and response of the proposer:

MODULE 2: **NEW SPECIES**

creating and naming one or more new species.

If more than one, they should be a group of related species belonging to the same genus. All new species must be placed in a higher taxon. This is usually a genus although it is also permissible for species to be “unassigned” within a subfamily or family. Wherever possible, provide sequence accession number(s) for **one** isolate of each new species proposed.

Code	2016.002aS	(assigned by ICTV officers)
To create 2 new species within:		
Genus:	<i>Avisivirus</i>	Fill in all that apply. • If the higher taxon has yet to be created (in a later module, below) write “ (new) ” after its proposed name. • If no genus is specified, enter “ unassigned ” in the genus box.
Subfamily:		
Family:	<i>Picornaviridae</i>	
Order:	<i>Picornavirales</i>	
Name of new species:	Representative isolate: (only 1 per species please)	GenBank sequence accession number(s)
<i>Avisivirus B</i>	ChPV-2 44C (chicken picornavirus 2)	KF979333
<i>Avisivirus C</i>	ChPV-3 45C (chicken picornavirus 3)	KF979334

Reasons to justify the creation and assignment of the new species:

- Explain how the proposed species differ(s) from all existing species.
 - If species demarcation criteria (see module 3) have previously been defined for the genus, **explain how the new species meet these criteria.**
 - If criteria for demarcating species need to be defined (because there will now be more than one species in the genus), please state the proposed criteria.
- Further material in support of this proposal may be presented in the Appendix, Module 11

Novel picornaviruses were detected in cloacal and tracheal samples of clinically healthy chickens in Hong Kong and one diarrheic chicken in Hungary. The novel picornaviruses share significant similarities to *Avisivirus A1*, i.e.

(i) the *Phasianidae* host,

(ii) similar genome layout (compare Appendix Figure 1):

VPg+5'UTR^{IRES-II}[1AB-1C-1D-2A1a^{NPG↓P}/(2A1b^{NPG↓P})/2A2^{NTPase}-2A3^{H-box/NC}-2B-2C^{Hel}/3A-3B^{VPg}-3C^{Prot}-3D^{Pol}]3'UTR-poly(A);

(iii) significant amino acid identities of capsid proteins (>40%) and 3CD (>50%) protein (compare **Tables 1, 2; Appendix**)

(iv) clustering with *Avisivirus A* in phylogenetic trees (compare **Figures 2,3; Appendix**).

The proposed *Avisivirus B1* has two NPG↓P motifs whereas the proposed *Avisivirus C1* has one.

MODULE 11: **APPENDIX**: supporting material

additional material in support of this proposal

References:

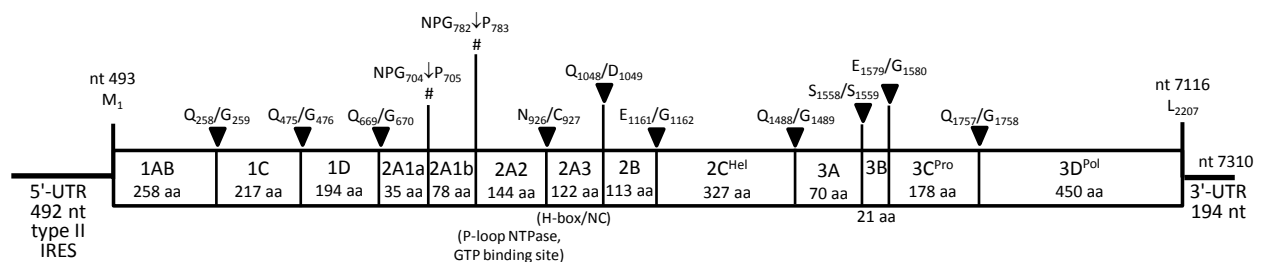
- Lau SKP, Woo PCY, Yip CCY, Li KSM, Fan RYY, Bai R, Huang Y, Chan KH, Yuen KY. 2014. Chickens host diverse picornaviruses originated from potential interspecies transmission with recombination. J. Gen. Virol. 95:1929-1944.
- Boros A, Pankovics P, Adonyi A, Fenyvesi H, Day JM, Phan TG, Delwart E, Reuter G. 2016. A diarrheic chicken simultaneously co-infected with multiple picornaviruses: Complete genome analysis of avian picornaviruses representing up to six genera. Virology 489:63-74.

Annex:

Include as much information as necessary to support the proposal, including diagrams comparing the old and new taxonomic orders. The use of Figures and Tables is strongly recommended but direct pasting of content from publications will require permission from the copyright holder together with appropriate acknowledgement as this proposal will be placed on a public web site. For phylogenetic analysis, try to provide a tree where branch length is related to genetic distance.

Genome organization:

Proposed: *Avisivirus B*, chicken picornavirus 2 [44C], GenBank acc. no. KF979333



Proposed: *Avisivirus C*, chicken picornavirus 3 [45C], GenBank acc. no. KF979334

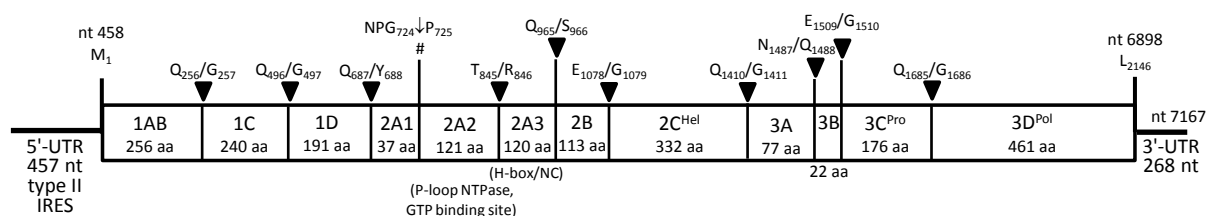


Figure 1: Schematic depiction of the avisivirus B and C genomes (*top*: chicken picornavirus 2 [44C], *below*: chicken picornavirus 3 [45C]). The open reading frames are indicated by boxes. Positions of putative nt and aa cleavage sites and the lengths of the deduced proteins are shown as proposed by Lau et al., 2014. Triangles (▼) indicate the putative 3C^{pro} cleavage sites; the hash (#) indicates ribosomal skipping sites at the NPG↓P motif. Note that *Avisivirus B* has two NPG↓P motifs whereas *Avisivirus C* has only one.

Fig. 2
P1

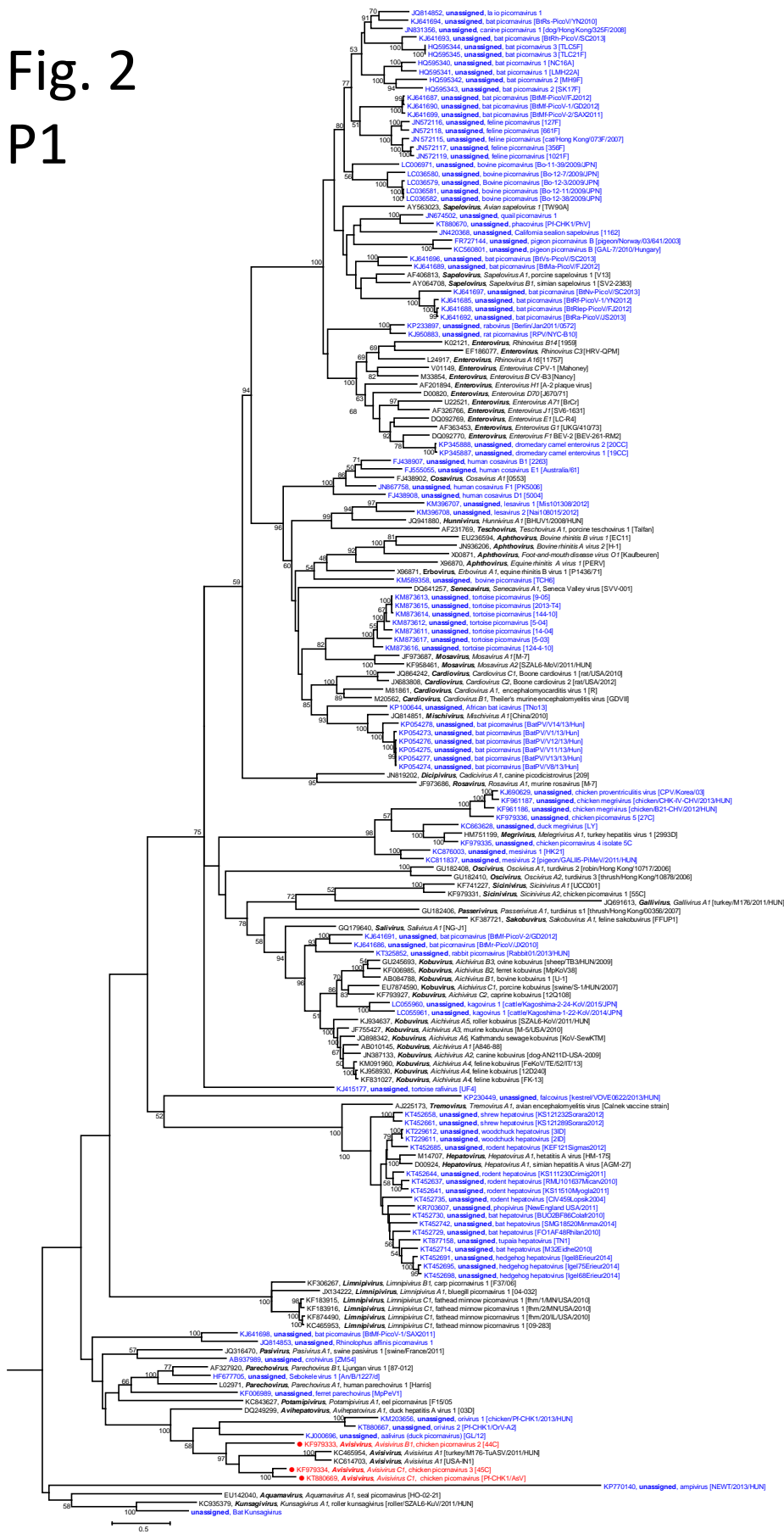


Figure 2: Phylogenetic analyses of picornavirus P1 using Maximum Likelihood tree inference (MEGA 5.2). 178 picornavirus sequences were retrieved from GenBank. Presented are GenBank accession numbers, *genus names*, *species names* and *types*. If available, common names and designations of isolates [in square brackets] are given. Yet unassigned viruses are printed in blue. Proposed names are printed in red and indicated by a dot (●). Numbers at nodes indicate bootstrap values obtained after 200 generations. The optimal substitution model (GTR+G+I) was determined with MEGA 5. The scale indicates substitutions/site.

Fig. 3
3CD

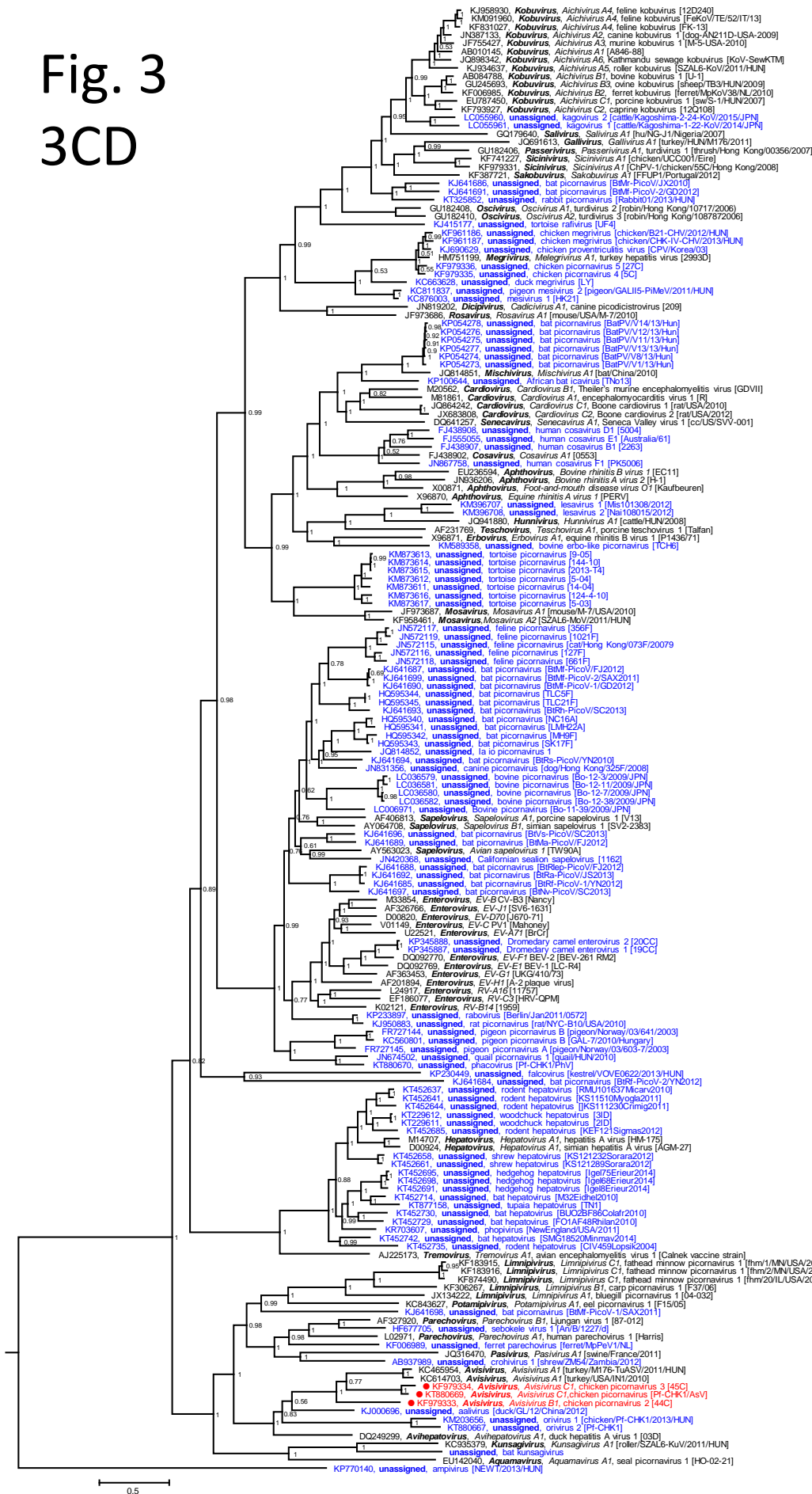


Figure 3: Phylogenetic analyses of picornavirus 3CD gene regions using Bayesian tree inference (MrBayes 3.2). 178 sequences were retrieved from GenBank. Presented are GenBank accession numbers, *genus names*, *species names* and *types*. If available, common names and designations of isolates [in square brackets] are given. Yet unassigned viruses are printed in blue. Proposed names are printed in red and indicated by a dot (●). Numbers at nodes indicate posterior probabilities obtained after 4,750,000 generations. The optimal substitution model (GTR+G+I) was determined with MEGA 5. The scale indicates substitutions/site.

Table 1. Estimates of Evolutionary Divergence between P1 Sequences

[1	2	3	4	5	6]
[1] KC465954 Avisivirus A1 [turkey/M176-TuASV/2011/HUN]	0.0000						
[2] KC614703 Avisivirus A1 [USA-IN1]	0.1065	0.0000					
[3] KF979333 Avisivirus B1 Chicken picornavirus 2 [44C]	0.5828	0.5748	0.0000				
[4] KF979334 Avisivirus C1 Chicken picornavirus 3 [45C]	0.5493	0.5523	0.5338	0.0000			
[5] KT880669 Avisivirus C1 [Pf-CHK1/AsV]	0.5741	0.5698	0.5605	0.1011	0.0000		
[6] DQ249299 Avihepato A1 DHAV-1 [03D]	0.6571	0.6609	0.6518	0.6252	0.6360	0.0000	

The number of amino acid differences per site from between sequences are shown. The analysis involved 6 amino acid sequences. The coding data was translated assuming a Standard genetic code table. All ambiguous positions were removed for each sequence pair. There were a total of 734 positions in the final dataset. Evolutionary analyses were conducted in MEGA5 [1].

Table 2. Estimates of Evolutionary Divergence between 3CD Sequences

[1	2	3	4	5	6]
[1] KC465954 Avisivirus A1 [turkey/M176-TuASV/2011/HUN]	0.0000						
[2] KC614703 Avisivirus A1 [turkey/USA/IN1/2010]	0.0409	0.0000					
[3] KF979333 Avisivirus B1 chicken_picornavirus_2_[44C]	0.4857	0.4848	0.0000				
[4] KF979334 Avisivirus C1 chicken_picornavirus_3_[45C]	0.4606	0.4597	0.4880	0.0000			
[5] KT880669 Avisivirus C1 [Pf-CHK1/AsV]	0.4558	0.4550	0.4896	0.0141	0.0000		
[6] DQ249299 Avihepatovirus DHAV-1_[03D]	0.6134	0.6208	0.6010	0.6045	0.6013	0.0000	

The number of amino acid differences per site from between sequences are shown. The analysis involved 6 amino acid sequences. The coding data was translated assuming a Standard genetic code table. All ambiguous positions were removed for each sequence pair. There were a total of 649 positions in the final dataset. Evolutionary analyses were conducted in MEGA5 [1].

P1:	intra-typic	observed aa divergence: <11%	⇒	aa identity: >89%
	inter-typic (within species)	-		
	between species	observed aa divergence: 53-58%	⇒	aa identity: >42%
	between genera	observed aa divergence: >62%	⇒	aa identity: <38%
3CD:	intra-typic	observed aa divergence: <5%	⇒	aa identity: >95%
	inter-typic (within species)	-		
	between species	observed aa divergence: 45-49%	⇒	aa identity: >51%
	between genera	observed aa divergence: >60%	⇒	aa identity: <40%